Detection of Ly-6B.2 Alloantigen in Formalin-Fixed, Paraffin-Embedded Mouse Tissue

Reagent and Antibody Information

1X Wash Buffer
3% Hydrogen Peroxide
1% BSA Diluent
Carezyme II (Pepsin)
DAB Chromogen
Hematoxylin

Blocking Serum: Normal Rabbit Serum

Vector Laboratories, Inc. Burlingame, CA 94010 www.vectorlabs.com 1-800-227-6666 Catalog# PK6104

Avidin / Biotin Blocking Kit

Vector Laboratories, Inc. Burlingame, CA 94010 www.vectorlabs.com 1-800-227-6666 Catalog # SP-2001

Primary Antibody: Rat Ly-6B.2 ALLOANTIGEN Monoclonal Antibody

AbD Serotec, Inc. Raleigh, NC 27604 1-919-878-7978 www.ab-direct.com Catalog # MCA771G Lot # 0911

Negative Control Serum: Purified Rat IgG2a Isotype Control Serum

BD Biosciences San Jose, CA 95131 www.bdbiosciences.com 1-855-236-2772 Catalog # 559073

Secondary Antibody: Biotinylated Rabbit Anti-Rat IgG (H+L)

Vector Laboratories, Inc. Burlingame, CA 94010 www.vectorlabs.com 1-800-227-6666 Catalog # BA-4001 Label Complex: R.T.U. Vectastain Elite ABC Reagent

Vector Laboratories, Inc. Burlingame, CA 94010 www.vectorlabs.com 1-800-227-6666 Catalog # PK-7100

Staining Procedure

Positive Control Tissue: Endothelial cells

Stain Localization: Cytoplasmic

1. Deparaffinize and hydrate slides through the following solutions:

Solution	Repetitions	Time
Xylene	2 times	5 minutes
100% Ethanol	2 times	3 minutes
95% Ethanol	2 times	3 minutes
1X Wash Buffer	2 times	5 minutes

- 2. Quench endogenous peroxidase by placing the slides in 3% hydrogen peroxide for 15 minutes.
- 3. Rinse the slides in 2 changes of 1X wash buffer for 5 minutes each.

4.	Proteolytic-Induced Epitope Retrieval Using Pepsin		
	ncubate the slides in Carezyme II: Pepsin (predilute) for 3 minutes at 37°C.		
	Allow the pepsin to reach room temperature before use.)		
	Rinse the slide in distilled water for 1 minute to stop the enzymatic reaction.		
5.	Rinse the slides in 2 changes of 1X wash buffer for 5 minutes each.		
6.	Block with 10% normal rabbit serum for 20 minutes at room temperature.		
	Lot # Date Reconstituted		
	OO NOT RINSE THE SLIDES. CONTINUE TO AVIDIN-BIOTIN BLOCK		
7.	Avidin / Biotin Blocking Kit		
	Lot # Exp. Date New Kit: yes / no		
	Apply avidin block for 15 minutes at room temperature.		

	Apply biotin block for 15 minutes at room temperature.
	DO NOT RINSE SLIDES WITH BUFFER BEFORE ADDING PRIMARY ANTIBODY. ONLY WIPE EXCESS BLOCK.
8.	Apply primary antibody at a 1:500 dilution. Incubate for 1 hour at room temperature. Lot # Exp. Date

For negative control slides, dilute rat IgG2a control serum so that it's IgG2a protein concentration matches that of the primary antibody (if necessary). Then make a 1:500 dilution. If the concentrations can't be matched using this method, the dilution for the negative reagent may need to be adjusted. Apply the negative and incubate for 1 hour at room temperature. Lot # Exp. Date		
9. Rinse the slides in 2 changes of 1X wash buffer for 5 minutes each.		
10. Apply the rabbit anti-rat secondary antibody at a 1:500 dilution. Incubate for 30 minutes at room temperature. Lot # Date Reconstituted		
11. Rinse the slides in 2 changes of 1X wash buffer for 5 minutes each.		
12. Apply the Vectastain R.T.U. Elite Label and incubate for 30 minutes at room temperature. Exp. Date New Kit: yes / no		
13. Rinse the slides in 2 changes of 1X wash buffer for 5 minutes each.		
14. Apply the DAB chromogen. Incubate in the dark for 6 minutes at room temperature. (Add 1 drop of DAB per ml of substrate) Lot # Exp. Date New Kit: yes / no		
15. Rinse the slides in tap water 3 minutes.		
16. Counterstain with hematoxylin for 20 seconds.		
7. Rinse the slides in tap water until water is clear.		
18. Gently agitate slides in 1X wash buffer until the tissues turn blue.		
19. Dehydrate through the following solutions:		

Solutions	Repetitions	Time
95% Ethanol	1 time	3 minutes
100% Ethanol	3 times	3 minutes
Xylene	2 times	5 minutes

20. Coverslip

Updated 02/21/13